Essentials of Glycobiology - A Survey

Dec 30 2019 - Jan 3, 2020 NCBS Graduate Elective Workshop Sessions 1-2, 10am-12noon and Session 3, 2-3pm

Course Director: Ajit Varki.

Distinguished Professor of Medicine and Cellular & Molecular Medicine Co-Director, Glycobiology Research and Training Center (<u>GRTC</u>) Co-Director, UCSD/Salk Center for Academic Research and Training in Anthropogeny (<u>CARTA</u>) <u>University of California, San Diego</u>, 9500 Gilman Drive, La Jolla, CA 92093-0687. Adjunct Professor, <u>Salk Institute for Biological Studies</u> Executive Editor, "<u>Essentials of Glycobiology</u>"

Every living cell in nature displays a dense and complex array of cell surface and secreted glycan chains, as well as intracellular glycosylation. For historical and technical reasons these essential components of life were poorly incorporated into the 20th century revolution in biology—becoming the "dark matter of the biological universe". With improved technologies and a flood of new information, the myriad roles of glycans in biology and pathology can now be fully integrated into our understanding of life. This course will provide and broad overview of the structure, evolution, biology, functions, pathology and practical applications regarding glycans in nature, using the text "*Essentials of Glycobiology*" *3rd edition, 2017*Cold Spring Harbor Laboratory Press. Full Text available <u>online</u> at NCBI, and a collection of slides presenting all figures can be found <u>here</u>. The online <u>Glycobiology GlossarySymbol Nomenclature</u> and <u>Study Guide</u> may be useful.

Students should have graduate level exposure to molecular and cellular biology. Sessions will consist of brief lectures and student presentations, with open discussion. Each Student will read 2 selected chapters in detail and be prepared to present a <u>10 min</u> overview of <u>ONLY</u> the key facts about chapter content, using online information and slides as needed. Grading based on attendance and participation.

Date/	Chapters	Presenter
Session		
Dec 30,	1. Historical Background and Overview2. Monosaccharide Diversity	AjitVarki
Session 1	3. Oligosaccharides and Polysaccharides4. Cellular Organization of Glycosylation	
	5. Glycosylation Precursors20. Evolution of Glycan Diversity	
	7. Biological Functions of Glycans	
Dec 30,	6. Glycosyltransferases and Glycan-Processing Enzymes	AnsumanBiswas
Session 2	9. N-Glycans	AnsumanBiswas
	<u>10. O-GalNAc Glycans</u>	KhushbooAgrawal
	11. Glycosphingolipids	RashmiGodbole
Dec 30,	12. Glycosylphosphatidylinositol Anchors	SarayuBeri
Session 3	13. Other Classes of Eukaryotic Glycans	MaroofHashmi
	14. Structures Common to Different Glycans	DivijKinger
	15. Sialic Acids and Other Nonulosonic Acids	ShahidHussain

Date/	Chapters	Presenter
Session		
Dec 31,	17. Proteoglycans and Sulfated Glycosaminoglycans	UtkarshAyyangar
Session 1	38. Proteins That Bind Sulfated Glycosaminoglycans	Arun GS
	<u>16. Hyaluronan</u>	Mohamed
	18. Nucleocytoplasmic Glycosylation	Poonam S
Dec 31,	19. The O-GlcNAc Modification	AdityaDeshpande
Session 2	21. Eubacteria	Shreyan Ray
	22. Archaea	GauravKansagara
	<u>23. Fungi</u>	NileshAghera
Dec 31,	24. Viridiplantae and Algae	ShyamiliGoutham
Session 3	25. Nematoda	Steffi Raju
	26. Arthropoda	Poonam S
	27. Deuterostomes	UtkarshAyyangar
Jan 1,	28. Discovery and Classification of Glycan-Binding Proteins	AbhikDutta
Session 1	29. Principles of Glycan Recognition	Arun GS
	30. Structural Biology of Glycan Recognition	Abel CherianVarkey
	<u>31. R-Type Lectins</u>	DigvijayLalwani
Jan 1,	<u>32. L-Type Lectins</u>	Prachi Joshi
Session 2	<u>33.P-Type Lectins</u>	DrisyaDileep
	34. C-Type Lectins	KavyaShetty
	<u>35. I-Type Lectins</u>	TriptiKharbanda
Jan 1,	36. Galectins	ShahidHussain
Session 3	37. Microbial Lectins: Hemagglutinins, Adhesins, and Toxins	ShailyaVerma
	42. Bacterial and Viral Infections	AdityaDeshpande
	43. Parasitic Infections	KavyaShetty
Jan 2,	39. Glycans in Glycoprotein Quality Control	RashmiGodbole
Session 1	41. Glycans in Systemic Physiology	DrisyaDileep
	40. Free Glycans as Signaling Molecules	Shreyan Ray
	8. A Genomic View of Glycobiology	BharathSaravanan
Jan 2,	44. Genetic Disorders of Glycan Degradation	Steffi Raju
Session2	45. Genetic Disorders of Glycosylation	Abel CherianVarkey
	46. Glycans in Acquired Human Diseases	TriptiKharbanda
	47. Glycosylation Changes in Cancer	ShyamiliGoutham
Jan 2	48. Glycan-Recognizing Probes as Tools	Prachi Joshi
Session 3	49. Glycosylation Mutants of Cultured Mammalian Cells	GauravKansagara

Date/	Chapters	Presenter
Session		
	50. Structural Analysis of Glycans	NileshAghera
	51. Glycomics and Glycoproteomics	AbhikDutta
Jan 3,	52. Glycoinformatics	ShailyaVerma
Session 1	53. Chemical Synthesis of Glycans and Glycoconjugates	DigvijayLalwani
	54. Chemoenzymatic Synthesis of Glycans and Glycoconjugates	MaroofHashmi
	55. Chemical Tools for Inhibiting Glycosylation	SarayuBeri
Jan 3,	56. Glycosylation Engineering	DivijKinger
Session 2	57. Glycans in Biotechnology and the Pharmaceutical Industry	Mohamed
	58. Glycans in Nanotechnology	BharathSaravanan
	59. Glycans in Bioenergy and Materials Science	KhushbooAgrawal
Jan 3,	60. Future Directions in Glycosciences	AjitVarki
Session 3	Course Recap and Final discussion.	
Also See UCSE) GRTC Slide Resource on Following pages	

UCSD GRTC Slide Resource

The figures available through the above links to the NCBI website are deliberately downsized in quality for on-screen presentation. For access to and permission to reproduce high-quality figures, contact <u>Cold Spring Harbor Laboratory Press</u>.

Chapter 1 Historical Background and Overview

- Figure 1.2 Open-chain and ring forms of glucose
- Figure 1.3 Schematic representation of the Thy-1 glycoprotein
- **Figure 1.4** Examples of electron micrographs of glycans coating cell surfaces
- Figure 1.5 Examples of symbols and conventions for drawing glycan structures
- Figure 1.6 Common classes of animal glycans
- Figure 1.7 Glycan-protein linkages reported in nature
- **Figure 1.8** Biosynthesis, use and turnover of a common monosaccharide

Chapter 2 Monosaccharide Diversity

- Figure 2.1 Structures of glyceraldehyde and dihydroxyacetone
- Figure 2.2 D- and L-glucopyranose in Fischer projection and chair conformation
- **Figure 2.3** Fischer projections for the acyclic forms of the D series of aldoses
- Figure 2.4 Common monosaccharides found in vertebrates
- Figure 2.5 Cyclization of acyclic D-glucose to form pyranose and furanose structures
- Figure 2.6 Conversion from Fischer to Haworth projection
- Figure 2.7 Chair conformations

Chapter 3 Oligosaccharides and Polysaccharides

- Figure 3.1 Examples of branched structures in N- and O-linked glycans
- Figure 3.2 Repeating units of cellulose and starch showing conformation determining torsion angles ϕ and ψ
- Figure 3.3 Structures of repeating units glycosaminoglycans and conformations of heparan sulfate monosaccharides
- Figure 3.4 Schematic representation of repeating units of bacterial polysaccharides

Chapter 4 Cellular Organization of Glycosylation

- Figure 4.1 Initiation and maturation of eukaryotic glycoconjugates in the ER–Golgi–plasma membrane pathway
- Figure 4.2 Topology and localization of Golgi Golgiglycosyltransferases and glycosidases

Chapter 5 Glycosylation Precursors

- Figure 5.1 Biosynthesis and interconversion of monosaccharides
- Figure 5.2 Biosynthesis of UDP-xylose and the branched sugar donor UDP-apiose from UDP-GlcA
- Figure 5.3 Conversion of activated sugar donors
- Figure 5.4 Nucleotide sugar transporters for PAPS and ATP in Golgi membranes of mammals yeast protozoa, and plants

Chapter 6 Glycosyltransferases and Glycan-Processing Enzymes

- Figure 6.1
 Strict acceptor substrate specificity of glycosyltransferases illustrated by B blood group transferase

 Figure 6.2
 Human chorionic gonadotropin recognition determinants used by glycoprotein hormone GalNAc transferase

 Figure 6.2
 Demain church are a trained eight/transferase, showing eight/transferase
- **Figure 6.3** Domain structure of a typical sialyltransferase, showing sialyl motifs shared by this family of enzymes **Figure 6.4** Ribbon diagrams of representative GT-A, GT-B, GT-C, and lysozyme-type fold glycosyltransferases

- Figure 6.5 Schematic representation of (A) inverting and (B) retaining catalytic mechanisms
- **Figure 6.6** Catalytic site of bovine β 1-4 galactosyltransferase

Chapter 7 Biological Functions of Glycans

 Figure 7.1
 General classification of the biological functions of glycans

Figure 7.2 Approaches for elucidating the biological functions of glycans

Chapter 8 A Genomic View of Glycobiology

Figure 8.1 Schematic examples of modular GTs (glycosyltransferases)

Chapter 9 N-Glycans

- Figure 9.1 Types of N-glycans
- Figure 9.2 Dolichol phosphate (Dol-P)
- Figure 9.3 Synthesis of Dolichol-P-P-GlcNAc2Man9Glc3
- Figure 9.4 Processing and maturation of an N-glycan
- Figure 9.5 Branching and core modification of complex N-glycans
- Figure 9.6 Typical complex N-glycans found on mature glycoproteins.

Chapter 10 O-GalNAc Glycans

- Figure 10.1 A simplified model of a large secreted mucin
- Figure 10.2 Biosynthesis of core 1 and 2 O-GalNAc glycans as described in the text. Green lines are protein
- Figure 10.3 Biosynthesis of core 3 and 4 O-GalNAc glycans as described in the text. Green lines are protein

Chapter 11 Glycosphingolipids

- Figure 11.1 Structures of representative glycosphingolipids (GSLs) and glycoglycerolipids
- Figure 11.2 Glycosphingolipid (GSL) neutral cores and their designations based on IUPAC Nomenclature
- Figure 11.3 Glycosphingolipids are synthesized by stepwise addition of sugars to ceramide and then to glycans

Chapter 12 Glycosylphosphatidylinositol Anchors

- Figure 12.1 General structure of GPI anchors
- Figure 12.2 Chemical and enzymatic reactions of glycosylphosphatidylinositol (GPI) anchors
- Figure 12.3 Glycosylphosphatidylinositol (GPI)-biosynthetic pathways of mammalian cells and Trypanosoma brucei
- Figure 12.4 Predicted topologies of the components of glycosylphosphatidylinositol (GPI) biosynthesis in mammalian cells
- Figure 12.5 Features of glycosylphosphatidylinositol (GPI)-anchored proteins and processing by GPI transamidase

Chapter 13 Other Classes of Eukaryotic Glycans

- Figure 13.1 Modifications of epidermal growth factor (EGF)-like repeats
- Figure 13.2 Extracellular domain of Notch showing the evolutionarily conserved sites for O-fucose and O-glucose
- Figure 13.3 Notch signaling pathway
- Figure 13.4 Modifications of thrombospondin type-1 repeats (TSRs)
- Figure 13.5 Biosynthetic pathway for O-mannose glycans
- Figure 13.6 Biosynthetic pathway for C-mannosylation and structural details of tryptophan-7 in RNase 2

Chapter 14 Structures Common to Different Glycans

Figure 14.1	N-Glycan synthesis generates complex N-glycans with branching GlcNAc residues that are
	usually extended
Figure 14.2	Terminal GlcNAc residues are usually galactosylated
Figure 14.3	Blood group i and I antigen synthesis
Figure 14.4	Type-1, -2, and -3 H, A, and B antigens that form the O (H), A, and B blood group determinants
Figure 14.5	Synthesis of H (O), A, and B blood group determinants
Figure 14.6	Type-1 and -2 Lewis determinants
Figure 14.7	Biosynthesis of antigens of the P1PK blood group system: Pk, P, and P1
Figure 14.8	Structure and synthesis of the Galα1-3Gal antigen
Figure 14.9	Structure and synthesis of N-glycans bearing terminal GalNAc, including those with sulfated-GalNAc
Figure 14.10	Synthesis of the human Sda or mouse CT antigen and the glycolipid GM2
Figure 14.11	Synthesis of α 2-6 and α 2-3 Sias on O-glycans and glycolipids by ST3Gal and ST6GalNAc
	sialyltransferases
Figure 14.12	Structure and synthesis of glycans with α 2-8 sialic acids, including PolySia on N-glycans
01	Otalia Astronom Lovins New Jackste Astro

- Chapter 15 Sialic Acids and Other Nonulosonic Acids
- Figure 15.1 Sialic acids (Sias) and other nonulosonic acids (NulOs)
- Figure 15.2 Diversity in sialic acid linkages
- Figure 15.3 Hierarchical levels of sialome complexity
- Figure 15.4 Metabolism of N-acetylneuraminic acid in vertebrate cells

Chapter 16 Hyaluronan

Figure 16.1	Hyaluronan consists of repeating disaccharides composed of N-acetylglucosamine (GlcNAc) and
	glucuronic acid (GIcA)
Figure 16.2	Hyaluronan biosynthesis by hyaluronan synthase
Figure 16.3	Modular organization of the link module superfamily of hyaluronan-binding proteins
Figure 16.4	The large cartilage chondroitin sulfate (CS) proteoglycan (aggrecan) forms an aggregate with
	hyaluronan and link protein
Figure 16.5	Structure of the link module
Figure 16.6	Hyaluronan capsule

Chapter 17 Proteoglycans and Sulfated Glycosaminoglycans

- Figure 17.1Proteoglycans consist of a protein core and one or more covalently attached glycosaminoglycan chainsFigure 17.2Glycosaminoglycans consist of repeating disaccharide unitsFigure 17.3Keratan sulfates are sulfated poly-N-acetyllactosamine chains -linked to either Asn or Ser/Thr residues
- Figure 17.4 Biosynthesis of chondroitin sulfate and HS initiated by the formation of a linkage region tetrasaccharide
- Figure 17.4 Biosynthesis of chondroitin sulfate/dermatan sulfate

Chapter 18 Nucleocytoplasmic Glycosylation

- Figure 18.1 Mechanism of glycosylation of Skp1 in the cytoplasm of protists
- Figure 18.2 Topology of glycosylation reactions and the destinations of their product glycoconjugates

Chapter 19 The O-GIcNAc Modification

- Figure 19.1 Many nuclear, mitochondrial, and cytoplasmic proteins are modified by O-linked β-GlcNAc (O-GlcNAc)
- Figure 19.2 (A) O-GlcNAcylated proteins occur in many different cellular compartments

- Figure 19.3 O-GlcNActransferase (OGT) is regulated by multiple complex mechanisms
- Figure 19.4 Elevating O-GlcNAc blocks insulin signaling at many points

Chapter 20 Evolution of Glycan Diversity

- Figure 20.1 Circular depiction of phylogeny of life on earth
- Figure 20.2 Dominant pathways of N-glycan processing among different eukaryotic taxa

Chapter 21 Eubacteria

- Figure 21.1 Organization of cell envelopes of Gram-negative bacteria, Gram-positive bacteria, and mycobacteria
- Figure 21.2 Structure, biosynthesis, and inhibition of peptidoglycan assembly
- Figure 21.3 Structures of additional cell wall polymers in classical Gram-positive bacteria and mycobacteria
- Figure 21.4 Structural organization of lipopolysaccharides (LPSs)
- Figure 21.5 Assembly and export of lipopolysaccharides
- Figure 21.6 Structures of exopolysaccharides and capsular polysaccharides

Chapter 22 Archaea

- Figure 22.1 Diversity of cell wall structure in the domain of Archaea
- Figure 22.2 The chemical structure of pseudomurein
- Figure 22.3 The structural diversity of N-linked glycans in Archaea
- Figure 22.4 The pathway of N-glycosylation in Haloferaxvolcanii

Chapter 23 Fungi

- Figure 23.1 Illustration of the cell wall of fungi, showing glycan polymers and mannoproteins
- Figure 23.2 Structures of selected yeast mannans
- Figure 23.3 Structures of selected O-linked glycans in fungi
- Figure 23.4 Biosynthesis of N-glycans and addition to -Asn-X-Ser/Thr- residues in newly synthesized glycoproteins
- Figure 23.5 Structures of two yeast glycosylphosphatidylinositol (GPI) anchors
- Figure 23.6 A quick-freeze deep-etch image of the edge of a Cryptococcus neoformanscell
- Figure 23.7 Structures of capsular polysaccharides in Cryptococcus neoformans

Chapter 24 Viridiplantae and Algae

- Figure 24.1 Glycosyl sequences of cellulose and selected hemicelluloses present in plant cell walls
- Figure 24.2 Schematic structure of pectin
- Figure 24.3 Schematic structure of proteoglycan referred to as arabinoxylan pectin arabinogalactan protein1 (APAP1)
- Figure 24.4 Types of N-glycans identified in plants
- Figure 24.5 Processing of N-glycans in the plant secretory system
- Figure 24.6 The most abundant plant galactolipids

Chapter 25 Nematoda

- Figure 25.1 Caenorhabditis elegans
- Figure 25.2 Life cycle of Caenorhabditis elegans
- Figure 25.3 Biosynthesis of paucimannosidic and core fucosylated N-glycans in Caenorhabditiselegans
- Figure 25.4 Biosynthesis of core-1 O-glycan in C. elegans and some O-glycans proposed to occur in adult worms
- Figure 25.5 Biosynthesis of chondroitin in Caenorhabditis elegans
- Figure 25.6 Chondroitin proteoglycans (CPGs) of Caenorhabditis elegans
- Figure 25.7 Examples of nematode glycolipids

Chapter 26 Arthropoda

- Figure 26.1 N-Linked glycan diversity in Drosophila and other insects
- Figure 26.2 Mutations in enzymes that process complex N-linked glycans alter brain morphology in D. melanogaster
- Figure 26.3 O-Linked glycan diversity in Drosophila and other insects
- Figure 26.4 Cell fate choices dependent on Notch require appropriate glycan expression
- Figure 26.5 Glycosaminoglycans regulate the contact-dependent maintenance of germline stem cells (GSCs)
- Figure 26.6 Glycosphingolipid glycan diversity

Chapter 27 Deuterostomes

- Figure 27.1 The purple sea urchin Strongylocentrotuspurpuratus. Sperm binding to a sea urchin egg
- Figure 27.2 Xenopuslaevis. Embryonic development from the neurula stage until just before hatching
- Figure 27.3 Zebrafish
- Figure 27.4 Cre-loxP targeting for making conditional gene knockouts in the mouse

Chapter 28 Discovery and Classification of Glycan-Binding Proteins

- Figure 28.1 Representative structures from four common animal lectin families
- Figure 28.2 Arrangements of carbohydrate-recognition domains (CRDs) in lectins
- Figure 28.3 Several major structural families of glycan-binding proteins (GBPs) and their biological distributions
- Figure 28.4 Mechanisms of enhanced binding of natural ligands to lectins

Chapter 29 Principles of Glycan Recognition

- Figure 29.1 Monovalent and multivalent interactions of a glycan-binding protein with glycan ligands
- Figure 29.2 Equations governing the interactions of a glycan-binding protein or lectin (L) with a glycan ligand (G)
- Figure 29.3 Frontal affinity chromatography, different glycan concentrations applied to a column of immobilized GBP
- **Figure 29.4** Example of isothermal titration calorimetry (ITC)
- **Figure 29.5** Example of surface plasmon resonance (SPR)
- Figure 29.6 Covalent glycan microarrays printed on N-hydroxysuccinimide (NHS)- or epoxide-activated glass slides

Chapter 30 Structural Biology of Glycan Recognition

- Figure 30.1 Representation of 6 calcium-dependent carbohydrate-binding sites found in crystal structures of lectins
- Figure 30.2 Distribution of lectins with structures in the 3D-Lectin database as a function of fold family
- Figure 30.3 Chemical shift mapping of exchange binding sites for a 4-sulfated CS hexamer on Link module of TSG6
- Figure 30.4 STD Binding epitope identification in complex-type glycan bound to HIV-1 neutralizing antibody PG16
- Figure 30.5 Docking of a heparan sulfate (HS) hexamer to the chemokine CXCL12a
- Figure 30.6 Interactions between donor acceptor and protein residues in the active site of ST6Gal1

Chapter 31 R-Type Lectins

- Figure 31.1 The R-type lectin superfamily
- Figure 31.2 Ricin and abrin
- **Figure 31.3** Structures of the β -trefoil R-type domains in different proteins
- Figure 31.4 Pathway of ricin uptake and toxic activity of the A chain in the cytoplasm results in cell death
- Figure 31.5 Structure and function of UDP-GalNAc

Chapter 32 L-Type Lectins

- Figure 32.1 Structure of ConA, a legume seed lectin in complex with a branched pentasaccharide
- Figure 32.2 Comparison of subunit structures of soybean agglutinin and human galectin-3 complexed glycan ligands

- **Figure 32.3** 3D structure of a peanut agglutinin (PNA) monomer showing the four loops involved in sugar binding
- Figure 32.4 Schematic representation of calnexin and its lectin domain, P domain and the calcium-binding domain

Chapter 33 P-Type Lectins

- Figure 33.1 Historical background regarding cross-correction of lysosomal enzyme deficiencies in cultured cells
- Figure 33.2 Pathways for biosynthesis of N-glycans bearing the mannose 6-phosphate (M6P) recognition marker
- **Figure 33.3** GlcNAc-P-T is an $\alpha 2\beta 2\gamma 2$ hexamer encoded by two genes
- Figure 33.4 Ribbon diagram of the bovine cation-dependent M6P receptor (CD-MPR)
- Figure 33.5 Subcellular trafficking pathways of glycoproteins, lysosomal enzymes, and M6P receptors (MPRs)

Chapter 34 C-Type Lectins

- Figure 34.1 Structure of C-type lectins (CTLs)
- Figure 34.2 Crystal structure of trimeric rat mannose-binding protein-A complexed with α-methylmannoside
- Figure 34.3 Different groups of C-type lectins (CTLs) and their domain structures
- Figure 34.4 Some C-type lectins (CTLs) are endocytic receptors
- Figure 34.5 Signaling activity of C-type lectins (CTLs) in innate immune responses
- Figure 34.6 Structures and functions of selectins

Chapter 35 I-Type Lectins

- Figure 35.1 Domain structures of the known Siglecs in humans and mice
- Figure 35.2 Structural basis of Siglec binding to ligands
- Figure 35.3 Proposed biological functions mediated by CD22
- Figure 35.4 Probable evolutionary chain of Red Queen effects involving Sias and CD33rSiglecs
- Figure 35.5 Proposed biological functions mediated by CD33-related Siglecs

Chapter 36 Galectins

- Figure 36.1 Different types of galectins in vertebrates and invertebrates and their organization and sequences
- Figure 36.2 (A) Ribbon diagram of the crystal structure of human galectin-1 complexed with lactose
- Figure 36.3 Structural aspects of galectins from mammals and invertebrates
- Figure 36.4 Possible biosynthetic routes for galectins in animal cells, using galectin-1 as an example
- Figure 36.5 Galectin interactions with cell-surface and extracellular ligands leads cell adhesion and signaling

Chapter 37 Microbial Lectins: Hemagglutinins, Adhesins, and Toxins

- Figure 37.1 Structure of the influenza virus hemagglutinin (HA) ectodomain
- Figure 37.2 Two views of a putative heparin sulfate-binding site on the dengue virus envelope protein
- Figure 37.3 Escherichia coli express hundreds of pili, indicated by the fine filaments extending from the bacterium
- Figure 37.4 The α anomer of mannose in the combining site of FimH
- Figure 37.5 Crystal structure of cholera toxin B-subunit pentamer bound to GM1 pentasaccharide

Chapter 38 Proteins That Bind Sulfated Glycosaminoglycans

- Figure 38.1 Conformation of heparin oligosaccharides
- Figure 38.2 Crystal structure of the antithrombin-pentasaccharide complex (from Protein Data Bank)
- Figure 38.3 Crystal and nuclear magnetic resonance (NMR) solution structures of GAG–protein complexes

Chapter 39 Glycans in Glycoprotein Quality Control

- Figure 39.1 Mature N-glycan
- Figure 39.2 Model of quality control in glycoprotein folding
- Figure 39.3 Degradation of oligomannosyl N-glycans in the ER, cytoplasm, and lysosomes

Chapter 40 Free Glycans as Signaling Molecules

- Figure 40.1 Plant defense response
- Figure 40.2 (A) Oligosaccharide elicitors of the plant defense response
- Figure 40.3 Generic structure of a Nod factor
- **Figure 40.4** A nonasaccharide from xyloglucan that shows signaling properties
- Figure 40.5 Schematic diagram of signaling pathways activated by binding of hyaluronan to CD44

Chapter 42 Bacterial and Viral Infections

- Figure 42.1 Classical experiments on the role of the pneumococcal polysaccharide capsule in virulence
- Figure 42.2 Activation of immune signaling by bacterial lipopolysaccharide (LPS)
- Figure 42.3 Examples of mechanisms of bacterial adherence to host-cell surfaces
- Figure 42.4 Structure of a polymicrobial biofilm
- Figure 42.5 Mechanisms of viral entry into host cells

Chapter 43 Parasitic Infections

- Figure 43.1 Life cycle of Plasmodium falciparum, a parasite that causes the most severe form of human malaria
- Figure 43.2 Schematic representation of the major surface glycoconjugates of procyclic and metacyclic Trypanosoma
- Figure 43.3 Schematic representation of the major surface glycoconjugates of Trypanosomacruzi
- Figure 43.4 Life cycle of *Leishmania* species
- Figure 43.5 Schematic representation of the major cell-surface glycoconjugates of Leishmania
- Figure 43.6 Structure of *Entamoebahistolytica* lipopeptidophosphoglycan (LPPG)
- Figure 43.7 Life cycle of Schistosoma species, the parasitic helminth that causes schistosomiasis in humans
- Figure 43.8 Glycan structures found in parasitic helminths, including Schistosomamansoni and Haemonchus contortus

Chapter 44 Genetic Disorders of Glycan Degradation

- Figure 44.1 Degradation of complex N-glycans
- Figure 44.2 Degradation of hyaluronan and heparan sulfate
- Figure 44.3 Degradation of chondroitin/dermatan sulfates (CS/DS) and keratan sulfate (KS)
- Figure 44.4 Degradation of glycosphingolipids

Chapter 45 Genetic Disorders of Glycosylation

- **Figure 45.1** Glycosylation-related disorders (graph)
- Figure 45.2 Congenital disorders of glycosylation in the N-glycosylation pathway
- Figure 45.3 UDP-Gal synthesis and galactosemia
- **Figure 45.4** O-Man glycan biosynthetic pathway

Chapter 47 Glycosylation Changes in Cancer

- Figure 47.1 N-Glycans increase in size on transformation, partly because of increased GlcNAc branching of N- glycans
- Figure 47.2 Loss of epithelial cell topology and polarization in cancer results in secretion of truncated mucins
- Figure 47.3 Gangliosides expressed in human neuroectodermal tumors

- Figure 47.4 Normal platelets, leukocytes and endothelial cells interact via selectins and selectin ligands
- Figure 47.5 Glycosaminoglycans (GAGs) in cancer

Chapter 48 Glycan-Recognizing Probes as Tools

- Figure 48.1 Examples of N-glycans recognized by concanavalin A (ConA) and Galanthusnivalis agglutinin (GNA)
- Figure 48.2 Examples of types of N-glycans recognized by L-PHA, E-PHA, and DSA
- Figure 48.3 Examples of types of glycan determinants bound with high affinity by different plant and animal lectins
- Figure 48.4 Examples of types of glycan determinants bound with high affinity by different plant lectins
- Figure 48.5 Examples of different mammalian glycan antigens recognized by specific monoclonal antibodies
- Figure 48.6 Additional examples of mammalian glycan antigens recognized by specific monoclonal antibodies
- Figure 48.7 Examples of different uses of plant and animal lectins, carbohydrate-binding molecules and antibodies
- Figure 48.8 Example of use of immobilized plant lectins in serial lectin affinity chromatography of glycopeptides

Chapter 49 Glycosylation Mutants of Cultured Mammalian Cells

- Figure 49.1 Alteration of cell-surface glycans by recessive and dominant glycosylation mutations
- Figure 49.2 Selections for glycosylation mutants
- Figure 49.3 Mutation of UDP-Gal-4-epimerase in IdID mutant CHO cells prevents generation of UDP-Gal and UDP-GalNAc

Chapter 50 Structural Analysis of Glycans

- Figure 50.1 Glycosidases used for structural analysis
- Figure 50.2 Example of linkage analysis showing a bacterial O-linked branched hexasaccharide
- Figure 50.3 Section of a nuclear magnetic resonance (NMR) spectrum of N-glycans released from the Fc of human IgG1
- **Figure 50.4** $\phi \psi$ energy plot for the glycosidic torsion angles between Glc residues in Glc β 1-4Glc-OMe

Chapter 51 Glycomics and Glycoproteomics

- Figure 51.1 Glycomics/glycoproteomics workflow for analysis of a purified glycoprotein
- Figure 51.2 Glycomics-assisted glycoproteomics of a complex mixture of glycoproteins
- Figure 51.3 Collision-induced dissociation-tandem mass spectrometry (CID-MS/MS) fragmentation of released N-glycans
- Figure 51.4 Complementary tandem mass spectrometry (MS/MS) fragmentation of N-glycopeptides

Chapter 52 Glycoinformatics

Figure 52.1 The critical role of glycomics in systems biology

Chapter 53 Chemical Synthesis of Glycans and Glycoconjugates

- **Figure 53.1** (A) Stereospecific formation of glycosidic bonds as either an α or β -linkage
- Figure 53.2 Protective group manipulations performed in one-pot procedures. Building blocks for glycan assembly
- Figure 53.3 Solution phase synthesis of Pseudomonas aeruginosa–derived decasaccharide 10
- Figure 53.4 Automated glycan assembly: (A) a tetrasaccharide and (B) an alginate dodecasaccharide
- Figure 53.5 cis-Arabinofuranosylation in the context of the synthesis of a plant cell wall arabinogalactan fragment 27

Chapter 54 Chemoenzymatic Synthesis of Glycans and Glycoconjugates

- Figure 54.1 Formation and hydrolysis of the glycosphingolipid, glucosylceramide
- Figure 54.2 Glycosyltransferase-mediated synthesis of sialyl Lewis x
- Figure 54.3 Glycosyltransferase-mediated synthesis of ganglio-oligosaccharides
- Figure 54.4 Chemoenzymatic synthesis of a library of mammalian N-glycans

- **Figure 54.5** (A) Equilibrium in a retaining β -glucosidase. (B) Mutant retaining β -glucosidase
- Figure 54.6 Glycosynthase-mediated synthesis of flavonoid glycosides
- Figure 54.7 A combined glycosyltransferase/glycosynthase/chemical synthesis of a lysosphingolipid
- Figure 54.8 Glycosynthase-mediated synthesis of homogeneous peptide N-glycans

Chapter 55 Chemical Tools for Inhibiting Glycosylation

- Figure 55.1 Structure of tunicamycin, which consists of uridine conjugated to the disaccharide, tunicamine
- Figure 55.2 Broad-spectrum inhibitors of the ppGalNAcTs identified from screening a uridine-based library
- Figure 55.3 Inhibitors of O-GlcNAc-specific β-hexosaminidase (OGA) and O-GlcNActransferase (OGT)
- Figure 55.4 Inhibitors of glycosphingolipid formation
- Figure 55.5 Structure of influenza neuraminidase inhibitors
- Figure 55.6 Crystallographic structures of influenza virus neuraminidases with inhibitors bound in the active site

Chapter 56 Glycosylation Engineering

- Figure 56.1 Overview of species-specific glycosylation features
- Figure 56.2 Precise gene editing modalities
- Figure 56.3 Complex N-glycan with glycosyltransferases responsible for each reaction

Chapter 57 Glycans in Biotechnology and the Pharmaceutical Industry

- Figure 57.1 Examples of carbohydrate-based drugs
- Figure 57.2 The synthetic influenza neuraminidase inhibitors Relenza and Tamiflu
- Figure 57.3 Glycomimetic E-selectin inhibitors based on sialyl Lewis x

Chapter 58 Glycans in Nanotechnology

- Figure 58.1 Different types of glyconanomaterials created by coupling glycans to surfaces of diverse nanomaterials
- Figure 58.2 Calculated representation of 2-nm-sized gold glyconanoparticle and corresponding TEM image
- Figure 58.3 (A) Binding studies using sLex-MNPs to rat E-selectin; (B) MRIs and 3D reconstruction of sLex-MNPs
- Figure 58.4 In vivo localization of filled-and-functionalized glyco-single-walled nanotubules (SWNTs)

Chapter 59 Glycans in Bioenergy and Materials Science

- Figure 59.1 Stacking of cellulose chains with regions of "order" and "disorder." Cellulose nanomaterial extraction
- Figure 59.2 Transmission electron microscopy images showing two types of cellulose nanomaterials